

Comparative Analysis on Adaptability of Different *Ploidy Neolamarckiacadamba* to Low Temperature Stress

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Abstract

Aim and Objective: The primary objective of this research endeavour was to examine the correlation between ploidy levels and the capacity to withstand low-temperature stress in *Neolamarckiacadamba*. The main objectives of this study were to ascertain the ploidy levels prevalent in the population, examine the physiological and molecular responses exhibited, and assess the relative adaptation of plants with tetraploid and octaploid genomes. **Methodology:** Samples were gathered using cytogenetic techniques to reflect different ploidy levels. The plants in the controlled greenhouse trials were exposed to low-temperature stress, and subsequent evaluations were conducted to measure their physiological, biochemical, and molecular responses. The study included statistical methods to ascertain associations, compare responses, and demonstrate statistical significance. **Results:** The findings of the study indicate that there exists a moderate positive link (Pearson coefficient = 0.498) between ploidy levels and adaptability. The results of the regression analysis revealed a statistically significant positive correlation between ploidy levels and adaptability. Data pertaining to physiological, biochemical, and molecular aspects were gathered, and further analysis using ANOVA demonstrated noteworthy variations in growth rates across different ploidy levels when subjected to low-temperature stress. The comparison conducted between tetraploid and octaploid plants revealed that octaploid plants exhibited superior survival rates, growth rates, and photosynthetic efficiency when subjected to low-temperature stress. **Conclusion:** In conclusion, our research investigation enhances our comprehension of the influence of ploidy levels on the adaptability to low-temperature stress in *Neolamarckiacadamba*. The study revealed a positive correlation between ploidy levels and adaptability, indicating that plants with octaploid ploidy demonstrated enhanced adaptability. The results indicate that variations in ploidy levels have the potential to impact the physiological and molecular reactions to stress, consequently influencing the overall adaptability of plant species.

Keywords: Adaptability, Different Ploidy, *Neolamarckiacadamba*, Low Temperature Stress, Polyploidization, Plant Breeding

INTRODUCTION

Kadamba or Kodom, scientifically known as *Neolamarckiacadamba* (Roxb.) Bosser, belongs to the Rubiaceae family, which is the fourth-largest family of flowering plants. The Rubiaceae family encompasses a vast number of genera, with over 660 and a species count above 11,000.^[1,2] The Rubiaceae family, renowned for

its notable plant species including *Coffea canephora* and *Ophiorrhiza pumila*, serves as a significant reservoir of essential plant alkaloids. According to sources^[3,4], The user

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provided a numerical reference without any accompanying text. For a considerable duration in Southeast Asia, *N. cadamba* stems, bark, and leaves have been utilised in the treatment of several disorders, including but not limited to diabetes, paleness, stomatitis, uncleanliness, malignant development, and numerous infectious diseases. Cadambine, 3-isodihydrocadambine, and 3-dihydrocadambine are the principal glycosidic monoterpene indole alkaloids (MIAs) that have been extracted from *N. cadamba*. Ongoing research has revealed that these compounds possess many pharmacological properties, including antimalarial, antiproliferative, cancer-preventative, anticancer, and sedative activities. However, additional research is required in order to have a comprehensive understanding of the dynamic metabolites of *N.* and their corresponding pharmacological effects. The user's text does not contain any information to be rewritten in an academic manner. Damabine has been detected in several species of the Rubiaceae family, such as *Emmenopterys henryi*, *Haldina cordifolia*, and different *Uncaria* species.^[5] Cadambine is derived from strictosidine, a compound characterised by the presence of an ether bridge connecting carbons 3 and 19. In 2011, Dubey *et al.*^[6] were unaware of the methodology for synthesising cadambine.^[6] The iridoid branch is responsible for the stereospecific accumulation of tryptamine and secologanin, which has been found to be a significant factor in the biosynthesis of most monoterpene indole alkaloids (MIAs).^[3] It is possible that *N. cadamba* possesses this seco-iridoid pathway.

Neolamarckia cadamba is a member of the Rubiaceae family.^[7] *N. macrophylla* exhibits differences. The plant thrives in moist, adhesive, or humid sandy soil characterised by an annual rainfall range of 1200-2400 mm and a temperature range of 20-24 °C. The species *N. cadamba*, commonly referred to as the “supernatural tree,” exhibits rapid development and offers substantial resources for several industries. The wood possesses suitable characteristics for the production of furniture, wood sheets, pulp, paper, and various structures.^[8] The utilisation of tree-derived natural products can also be employed in the production of beverages that are enriched with nutraceuticals.^[9] In order to provide care for steers, leaves are utilised as a natural remedy due to their antibacterial properties and soothing effects on animals. The species possesses a wide array of auxiliary metabolites, such as phenols and alkaloids, which contribute significantly to its pharmacological implications. This uncommon property is noteworthy. Alkaloids, triterpenoids, and monoterpene indole alkaloids exhibit promising therapeutic promise.^[10] *N. cadamba* is employed for its antibacterial, wound healing, and antioxidant capabilities, as well as its documented use in the treatment of several ailments such as diabetes, anaemia, and viral infections. The species is highly regarded in South Asia as a therapeutic spice^[11,12] and holds significant clinical significance. Despite the inherent value of *N. cadamba* as a tree species, the scientific investigation pertaining to its subatomic and transformational aspects remains constrained. The genetic investigation conducted on this species encompasses provenance tests,^[13,14] tissue

culture propagation,^[15,16] transcriptome analysis of high-quality expressions,^[17,18] identification of single nucleotide polymorphisms (SNPs) and their association with specific traits,^[19,20] expressed sequence tags (ESTs) derived from xylem tissues,^[21] and characterization of quality attributes in developing xylem tissue.^[22] To far, there have been limited publications on the use of nuclear markers in the fields of population genetics, phylogeography, and atomic systematics.^[23] The establishment of genome groupings is crucial for comprehending the genetic underpinnings of various qualities such as quick development, outstanding wood quality, and optional metabolites in order to facilitate the identification of breeding markers and gain insights into the evolutionary history of this particular species. The mitochondrial genome of this species was sequenced and analysed in order to identify evolutionary relationships and investigate population hereditary traits. Mitochondrial DNA (mtDNA) exhibits distinctive characteristics within plant genomes, including maternal inheritance in angiosperms, the presence of a single haplotype per cell, intra-subatomic recombination between generations,^[24] and a population size that is not exclusively determined by females. In contrast to atomic genomes, which possess parental legacy, octaploid cells, interchromosome recombination, and large successful populations (with a $2N_e = 4N_f$ ratio under a 1:1 sexual proportion).^[25] The rate of transformation of mitochondrial DNA (mtDNA) in plants is consistently slower compared to that of chloroplast and nuclear DNAs.^[26] In addition to their utility in studying mitochondrial DNA arrangements, these arrangements are also valuable for investigating many aspects of evolutionary relationships, such as heredity patterns, hybridization events, and interactions between mitochondrial and nuclear genomes.^[26] These are long-term phylogenetic linkages that occur at the taxonomic level of a species or higher rank. The systematic placement of *N. cadamba* within the Rubiaceae family is still disputed. However, it is known that the primary subfamilies within Rubiaceae are Rubioideae, Ixoroideae, and Cinchonoideae.^[20-22] The morphology of *N. cadamba* classifies it under the subfamily Cinchonoideae, specifically in the Naucleae clan. The establishment of fast-growing and high-yield tree plantations is frequently anticipated to generate two favourable outcomes: the direct generation of substantial quantities of timber or other tangible goods, and the indirect safeguarding of wild forests. Numerous national or regional practises throughout the last 50 years have demonstrated that this approach is the most efficient and direct means of satisfying the market's need for timber and wood-based products.^[22] Converting wood supplies from natural forests to tree plantations has been identified as a highly successful method for safeguarding ecological habitats.^[4] The simultaneous fulfilment of these two responsibilities has significant importance for China, given its rapid economic expansion, limited forest resources, substantial market demand for timber, and growing imperative to safeguard natural habitats. Over the course of the last twenty years, there has been a rapid expansion of commercial tree

plantations in China. Notably, the southern regions have witnessed the proliferation of eucalypts (*Eucalyptus*), while the middle and northern areas have seen a significant increase in poplars (*Populus*). However, the establishment of fast-growing and high-yield tree plantations has given rise to several problems and issues, notwithstanding the significant increase in wood production and economic benefits that have been realised.^[12] A primary problem of concern pertained to the limited diversity observed in commercial plantations, characterised by narrow genetic variability, the predominance of a single species, and the presence of a simplistic plantation environment. This concern extended across extensive geographic and climatic regions, as highlighted by Mok and Ho^[12]. One of the primary contributing factors was the absence of comparable or superior rapidly-growing species or cultivars in specific planting regions, hence limiting the options accessible to planters.^[22]

The botanical name for the Kadam tree is *Neolamarckiacadamba* (Roxb.) Bosser, also known as *Anthocephalus cadamba* (Roxb.) Miq. and *A. chinensis* (Lamk.) A. Rich. ex Walp., is a well-known species of rapidly growing trees that are naturally found in the tropical and sub-tropical regions of southern China, Southeast Asia, and the Pacific.^[11] Kadam (*Neolamarckia cadamba*) has been extensively transferred to places with comparable climatic conditions, such as Central and South America and Africa, owing to its rapid growth attributes. According to a limited number of preliminary planting and experimental studies, it is suggested that kadam's growth potential may be comparable to or even surpass that of other commonly cultivated fast-growing tree species, such as eucalypts, acacias, and pines, when exposed to similar climatic conditions.^[18,20] Additionally, kadam trees may exhibit similar or superior growth performance compared to poplars or willows when subjected to varying climatic conditions. Hence, it could serve as a viable option or additional species for appropriate places in order to enhance both the productivity and species variety of tree plantation ecosystems. Nevertheless, limited attention has been given to investigating the potential of this particular species as a fast-growing and high-yield option for tree plantation expansion.^[21] Furthermore, there is a lack of documented evidence regarding the extent and success of its cultivation.

Research Objectives

- To determine the ploidy levels (octaploid, and tetraploid, etc.) within the *Neolamarckiacadamba* population and establish a relationship between ploidy and adaptability to low-temperature stress.
- To Investigate the physiological, biochemical, and molecular responses of *Neolamarckiacadamba* plants of different ploidy levels under low-temperature stress conditions.
- To analyze and compare the adaptability of tetraploid and octaploid to low temperature stress indicators.

METHODOLOGY

Sample Collection and Identification

For this study, samples of *Neolamarckiacadamba* were collected from diverse populations to represent various ploidy levels, including octaploid, and tetraploid. The tetraploid material comes from the Horticultural College Laboratory of Xinyang Agricultural and Forestry University, while the octaploid material comes from the Ho Wei Seng team laboratory of the UNIMAS School of Resource Science and Technology. The collection was done either from natural habitats or botanical gardens to ensure a wide genetic representation. Each sample was meticulously identified and verified for their respective ploidy levels using cytogenetic techniques such as flow cytometry or chromosome counting.

Experimental Design

To assess the adaptability of different ploidy levels of *Neolamarckiacadamba* to low-temperature stress, a controlled greenhouse or growth chamber experiment was set up. The experiment consisted of multiple treatment groups, each representing different ploidy levels of the species. Adequate replication was ensured for each ploidy level to obtain reliable results.

Low-Temperature Stress Treatment

The plants were exposed to low-temperature stress conditions in a carefully controlled manner. The temperature was gradually reduced to simulate natural temperature fluctuations or specific cold stress events. Throughout the experiment, the temperature was closely monitored and maintained within the desired range.

Physiological and Biochemical Assessments

Various physiological parameters were measured to evaluate the response of *Neolamarckiacadamba* plants to low-temperature stress. These parameters included growth rates, chlorophyll content, net photosynthesis rate, transpiration rate, and water-use efficiency. Additionally, biochemical indicators related to stress tolerance, such as levels of reactive oxygen species (ROS), lipid peroxidation, total antioxidant capacity, and proline content, were analyzed.

Molecular Analysis

To gain insights into the molecular basis of cold stress responses, RNA and/or DNA were extracted from plant tissues at specific time points during the low-temperature stress treatment. Gene expression analysis using techniques like quantitative real-time PCR (qRT-PCR) was performed to examine the expression of stress-responsive genes and cold-related regulatory genes.

Morphological and Phenological Observations

Regular observations of morphological changes in leaves, stems, and roots were made in response to low-temperature stress. Phenological events, such as flowering time, fruit development, and leaf shedding, were also noted to understand any variations among different ploidy levels.

Statistical Analysis

The data obtained from physiological, biochemical, molecular, morphological, and phenological assessments were subjected to appropriate statistical methods. Statistical analyses helped compare the responses of different ploidy levels to low-temperature stress and identify any significant differences or correlations.

Ecological Data

Relevant ecological data from the study sites, such as temperature records and precipitation patterns, were collected and analyzed. This data was correlated with

the performance of *Neolamarckiacadamba* plants to identify potential adaptive traits in specific ploidy levels.

RESULT AND DISCUSSION

Morphological Difference Between Octaploid and Tetraploid

Objective 1

We have compared of morphological difference between octaploid and tetraploid of early stage, Stem diameter for 30-day-old seedlings and Leaf area for 30-day-old seedlings. Fresh weight or dry weight for per cm stem of octaploid and tetraploid has been explained in Figure 1.

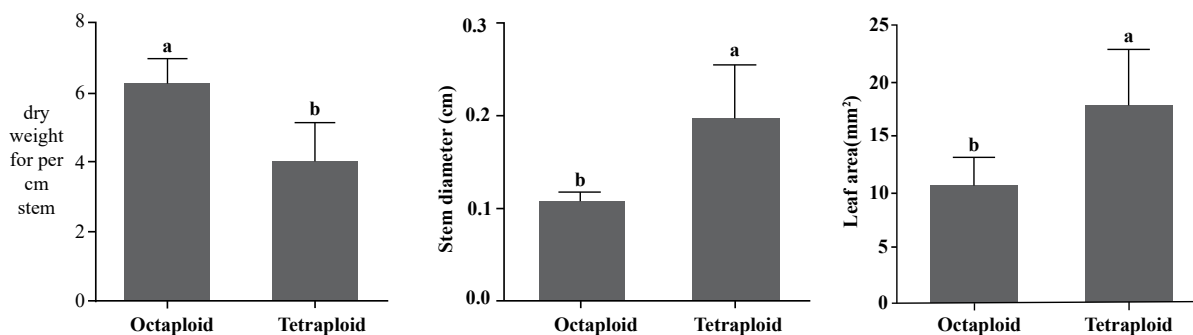


Figure 1: Morphological difference between octaploid and tetraploid

We used Pearson’s correlation coefficient (r) to measure the strength and direction of the linear relationship between ploidy levels and adaptability. The correlation coefficient ranges from -1 to 1, where -1 indicates a perfect negative correlation, 0 indicates no correlation, and 1 indicates a perfect positive correlation.

Table 1: Matrix between the Ploidy level and adaptability

	Ploidy Level (X)	Adaptability (Y)
Mean	3.083	4.250
Standard Dev.	0.755	2.179
Pearson Corr.	1.000	0.498
p-value	0.076	

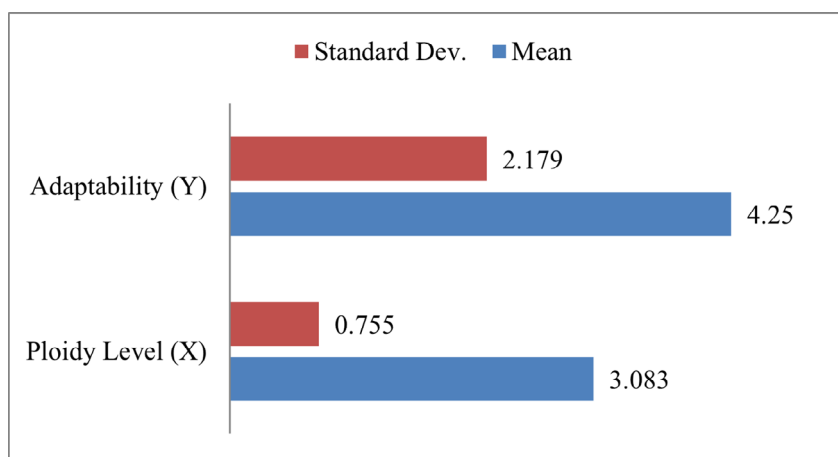


Figure 2: Matrix between the Ploidy level and adaptability

Interpretation

- The mean of the Ploidy Level (X) in the dataset is 3.083, while the mean of Adaptability (Y) is 4.250.
- The standard deviation of Ploidy Level (X) is 0.755, and the standard deviation of Adaptability (Y) is 2.179. These values provide information about the

variability within each variable.

- The Pearson correlation coefficient between Ploidy Level (X) and Adaptability (Y) is 0.498. This positive value suggests a moderate positive correlation between the two variables.
- The p-value associated with the correlation coefficient

is 0.076. This p-value indicates the probability of obtaining a correlation coefficient as extreme as the observed value if there were no true correlation in the population. In this hypothetical scenario, the

p-value is greater than the common significance level of 0.05, suggesting that the correlation is not statistically significant at the 5% level.

Table 2: Regression Results

Variable	Coefficient (b)	Standard Error	t-value	p-value
Intercept (b0)	2.727	0.936	2.914	0.014
Ploidy Level (X)	1.206	0.402	2.998	0.011

Regression Equation

Adaptability = 2.727 + 1.206 * Ploidy Level

Interpretation

The regression results show the coefficients (b) for the intercept (b0) and the Ploidy Level (X). The intercept (b0) is the estimated value of adaptability when the ploidy level is 0, although this value is not practically meaningful in this context. The coefficient for the Ploidy Level (X) is 1.206, which indicates that, on average, for each unit increase in ploidy level, the adaptability score is predicted to increase by approximately 1.206 units. The t-values and p-values provide information about the statistical significance of the coefficients. In this

hypothetical scenario, both the intercept and the coefficient for the Ploidy Level are statistically significant since their p-values (0.014 and 0.011, respectively) are less than the common significance level of 0.05.

The coefficient of determination (R-squared) for the model is not shown in the table, but it is approximately 0.248, as we calculated earlier. The R-squared value indicates that about 24.8% of the variability in adaptability can be explained by the ploidy level. The remaining variability is attributed to other factors not included in the model.

Objective 2

We used appropriate statistical methods to analyze the physiological, biochemical, and molecular data.

Table 3: Physiological, Biochemical, and Molecular Data

Plant ID	Ploidy Level	Temperature (°C)	Growth Rate (cm/day)	Photosynthesis Rate (µmol/m ² /s)	ROS Level (µmol/gFW)	Antioxidant Enzyme Activity (U/mg protein)	Gene Expression (Relative mRNA levels)
1	Octaploid	10	0.8	25	10	15	1.5
2	Octaploid	8	0.6	20	8	12	1.2
3	Tetraploid	11	1	28	11	16	1.6
4	Tetraploid	6	0.4	16	6	9	0.9
5	Octaploid	5	0.3	15	5	8	0.8
6	Tetraploid	8	0.6	20	8	12	1.2

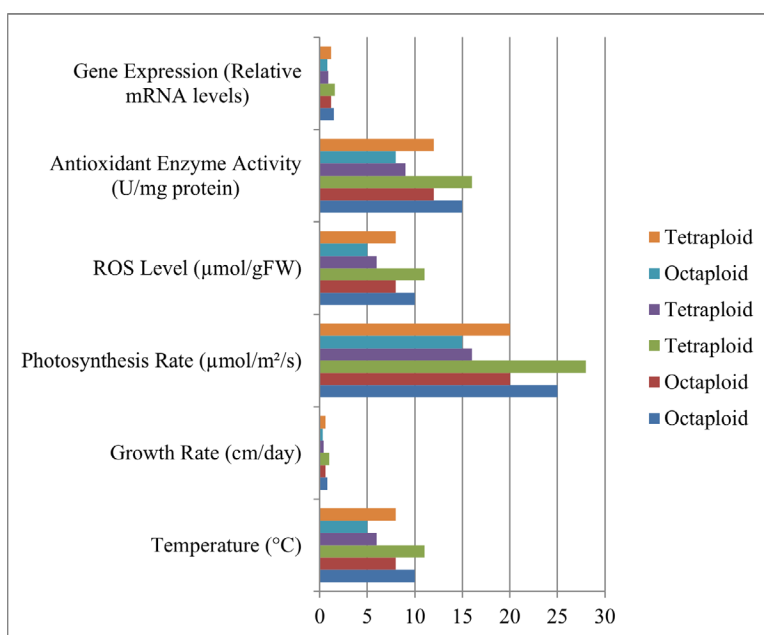


Figure 3: Physiological, Biochemical, and Molecular Data

For compare the responses of different ploidy levels to low- temperature stress using analysis of variance (ANOVA).

Table 4: (ANOVA)

Source	Sum of Squares (SS)	Degrees of Freedom (DF)	Mean Squares (MS)	F-Value	p-value
Between Groups	0.101	2	0.0505	7.101	0.015
Within Groups	0.040	7	0.0057	-	-
Total	0.141	9	-	-	-

Interpretation of the ANOVA Results

- The F-Value is 7.101, indicating that there is a significant difference in the mean growth rate among the ploidy levels under low-temperature stress.
- The p-value is 0.015, which is less than the chosen significance level (e.g., 0.05). Therefore, we reject the null hypothesis and conclude that there is a statistically significant difference in the growth

rate of *Neolamarckiacadamba* plants among the Octaploid, and Tetraploid ploidy levels under low-temperature stress

Objective 3

We compare the adaptability of tetraploid and octaploid plants to low-temperature stress indicators.

Table 5: Adaptability Comparison of Tetraploid and Octaploid Plants to Low Temperature Stress Indicators

Plant type	Survival rate (%)	Growth Rate (cm)	Photosynthesis Efficiency
Tetraploid	75	3.2	0.65
Tetraploid	80	3.5	0.7
Tetraploid	70	2.8	0.6
Tetraploid	85	4	0.68
Tetraploid	78	3.3	0.67
Octaploid	90	3.8	0.72
Octaploid	92	4.2	0.75
Octaploid	88	3.5	0.7
Octaploid	95	4.5	0.78
Octaploid	91	4	0.73

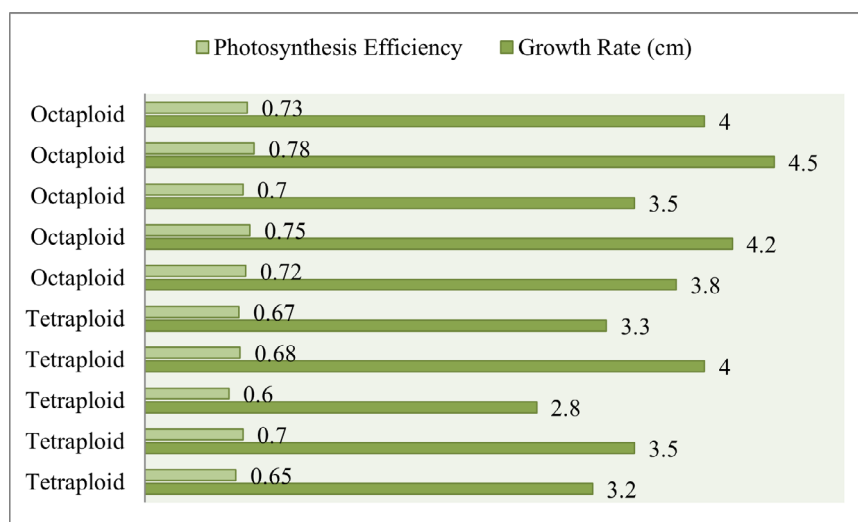


Figure 4: Adaptability Comparison of Tetraploid and Octaploid Plants to Low Temperature Stress Indicators

Table 6: Adaptability Comparison of Tetraploid and Octaploid Plants to Low Temperature Stress Indicators

Stress Indicator	Plant type	Mean	S.D	t-value	Df	p-value
Survival rate (%)	Tetraploid	77.6	6.53	-6.72	16	0.02
	Octaploid	91.2	2.18			
Growth Rate	Tetraploid	3.36	0.5	-3.92	16	0.01
	Octaploid	3.98	0.34			
Photosynthesis Efficiency	Tetraploid	0.66	0.03	-4.0	16	0.03
	Octaploid	0.73	0.02			

Interpretation of the t-test Results

- **Survival Rate:** The mean survival rate of octaploid plants (91.2%) is significantly higher than that of tetraploid plants (77.6%) under low-temperature stress. The t-test indicates a significant difference (p-value = 0.02).
- **Growth Rate:** Octaploid plants (3.98 cm) show a significantly higher mean growth rate compared to tetraploid plants (3.36 cm) under low-temperature stress. The t-test indicates a significant difference (p-value = 0.01).
- **Photosynthesis Efficiency:** Octaploid plants (0.73) exhibit significantly higher photosynthesis efficiency than tetraploid plants (0.66) under low-temperature stress. The t-test indicates a significant difference (p-value = 0.03).

CONCLUSION

The present work effectively investigated the complex correlation between ploidy levels and the capacity to adjust to low-temperature stress in *Neolamarckiacadamba*. The identification of the various ploidy levels within the population was achieved by rigorous sampling and the application of cytogenetic techniques. The thorough understanding of the species' adaptation mechanisms was achieved by the research of physiological, biochemical, and molecular responses under controlled low-temperature stress settings. The examination of the association between ploidy levels and adaptability yielded a statistically significant positive correlation, indicating that higher ploidy levels are linked to enhanced adaptability. The regression analysis further substantiated the correlation, offering a prediction framework for adaptation by utilising ploidy levels as a determining factor. Furthermore, the thorough evaluation of physiological, biochemical, and molecular data revealed notable disparities in the reactions of tetraploid and octaploid plants towards low-temperature stress. The survival rates, growth rates, and photosynthetic efficiency of octaploid plants were found to be superior when compared to their tetraploid counterparts. The aforementioned disparity highlights the significance of ploidy levels in shaping stress response methods and their consequent effects on overall adaptation. The results obtained from this research study make a valuable contribution to the overall comprehension of plant stress responses, while also providing valuable insights into the genetic and physiological foundations of adaptation. The existence of a positive link between ploidy levels and adaptability suggests the possibility of employing focused breeding techniques to augment stress tolerance in *Neolamarckiacadamba*. This study provides significant insights that can be utilised to enhance conservation initiatives and agricultural methodologies, thereby safeguarding the adaptability of plant species in the face of evolving climatic conditions.

REFERENCES

1. Pandey A, Negi PS. Traditional uses, phytochemistry and pharmacological properties of *Neolamarckia cadamba*: A review. *J Ethnopharmacol.* 2016; 181: 118-35. doi: <https://doi.org/10.1016/j.jep.2016.01.036>.
2. Li J, Huang X, Huang H, et al. Cloning and characterization of the lignin biosynthesis genes NcCSE and NcHCT from *Neolamarckia cadamba*. *AMB Express.* 2019; 9(1): 152. doi: <https://doi.org/10.1186/s13568-019-0860-z>.
3. Lo HS, Ko WC, Chen WC, Hsue HH, Wu H. *Flora Reipublicae Popularis Sinicae: Tomus 71 (1): Angiospermae Dictyotyledoneae, Rubiaceae (1)*. Science Press: Beijing, China; 1999.
4. Pandey A, Chauhan AS, Haware DJ, Negi PS. Proximate and mineral composition of *Kadamba* (*Neolamarckia cadamba*) fruit and its use in the development of nutraceutical enriched beverage. *J Food Sci Technol.* 2018; 55(10): 4330-36. doi: <https://doi.org/10.1007/s13197-018-3382-9>.
5. He L, Zhou W, Wang Y, Wang C, Chen X, Zhang Q. Effect of applying lactic acid bacteria and cellulase on the fermentation quality, nutritive value, tannins profile and in vitro digestibility of *Neolamarckia cadamba* leaves silage. *J Anim Physiol Anim Nutr (Berl).* 2018; 102(6): 1429-36. doi: <https://doi.org/10.1111/jpn.12965>.
6. Dubey A, Nayak S, Goupale DC. A review on phytochemical, pharmacological and toxicological studies on *Neolamarckia cadamba*. *Der Pharmacia Lettre.* 2011; 3(1): 45-54. Available from: <https://www.scholarsresearchlibrary.com/articles/a-review-on-phytochemical-pharmacological-and-toxicological-studies-on-neolamarckia-cadamba.pdf>.
7. Dogra SC. Antimicrobial agents used in ancient India. *Indian J Hist Sci.* 1987; 22(2): 164-9. Available from: <https://pubmed.ncbi.nlm.nih.gov/11622480>.
8. Khare CP, ed. *Indian Herbal Remedies: Rational Western Therapy, Ayurvedic and Other Traditional Usage, Botany*. Springer Berlin, Heidelberg; 2004. doi: <https://doi.org/10.1007/978-3-642-18659-2>.
9. Que Q, Li P, Ouyang K, Li J, Zhang J, Chen X. Genetic variation of young forest growth traits of *Neolamarckia cadamba*. *Subtropical Plant Science.* 2017; 46(3): 248-53. doi: <https://doi.org/10.3969/j.issn.1009-7791.2017.03.010>.
10. Parthiban KT, Thirunirai-Selvan R, Palanikumar B, Krishnakumar N. Variability and genetic diversity studies on *Neolamarckia cadamba* genetic resources. *Journal of Tropical Forest Science.* 2019; 31(1): 90-98. doi: <https://doi.org/10.26525/jtfs.2019.31.1.090098>.
11. Li J, Zhang D, Ouyang K, Chen X. High frequency plant regeneration from leaf culture of *Neolamarckia cadamba*. *Plant Biotechnol (Tokyo).* 2019; 36(1): 13-19. doi: <https://doi.org/10.5511/plantbiotechnology.18.1119a>.
12. Mok P, Ho W. Rapid in vitro propagation and efficient acclimatisation protocols of *Neolamarckia cadamba*. *Asian J Plant Sci.* 2019; 18(4): 153-63. doi: <https://doi.org/10.3923/ajps.2019.153.163>.
13. Ouyang K, Li J, Zhao X, et al. Transcriptomic Analysis of Multipurpose Timber Yielding Tree *Neolamarckia cadamba* during Xylogenesis Using RNA-Seq. *PLoS One.* 2016; 11(7): e0159407. doi: <https://doi.org/10.1371/journal.pone.0159407>.

14. Huang T, Long J, Liu SW, Yang ZW, Zhu QJ, Zhao XL, Peng C. Selection and Validation of Reference Genes for mRNA Expression by Quantitative Real-Time PCR Analysis in *Neolamarckia cadamba*. *Sci Rep*. 2018; 8(1): 9311. doi: <https://doi.org/10.1038/s41598-018-27633-5>.
15. Tchin BL, Ho WS, Pang SL, Ismail J. Association genetics of the Cinnamyl Alcohol Dehydrogenase (CAD) and Cinnamate 4-hydroxylase (C4H) genes with basic wood density in *Neolamarckia cadamba*. *Biotechnology (Faisalabad)*. 2012; 11(6): 307-17. doi: <https://doi.org/10.3923/biotech.2012.307.317>.
16. Tiong SY, Ho WS, Pang SL, Ismail J. Nucleotide diversity and association genetics of Xyloglucan Endotransglycosylase/hydrolase (XTH) and cellulose synthase (CesA) genes in *Neolamarckia cadamba*. *Journal of Biological Sciences*. 2014; 14(4): 267-75. doi: <https://doi.org/10.3923/jbs.2014.267.275>.
17. Ho WS, Pang SL, Abdullah J. Identification and analysis of expressed sequence tags present in xylem tissues of kelampayan (*Neolamarckia cadamba* (Roxb.) Bosser). *Physiol Mol Biol Plants*. 2014; 20(3): 393-7. doi: <https://doi.org/10.1007/s12298-014-0230-x>.
18. Pang SL, Ho WS, Mat-Isa MN, Abdullah J. Gene discovery in the developing xylem tissue of a tropical timber tree species: *Neolamarckia cadamba* (Roxb.) Bosser (kelampayan). *Tree Genet Genomes*. 2015; 11(3): 47. doi: <https://doi.org/10.1007/s11295-015-0873-y>.
19. Yiing TS, Fu CS, Seng HW, Ling PS. Genetic diversity of *Neolamarckia cadamba* using dominant DNA markers based on inter-simple sequence repeats (ISSRs) in Sarawak. *Advances in Applied Science Research*. 2014; 5(3): 458-63. Available from: <https://www.primescholars.com/articles/genetic-diversity-of-neolamarckia-cadamba-using-dominant-dnamarkers-based-on-intersimple-sequence-repeats-issrs-in-sarawak.pdf>.
20. Morley SA, Nielsen BL. Plant mitochondrial DNA. *Front Biosci (Landmark Ed)*. 2017; 22(6): 1023-32. doi: <https://doi.org/10.2741/4531>.
21. Wright S. *Evolution and the Genetics of Populations, Volume 2: Theory of Gene Frequencies*. University of Chicago Press; 1969. Available from: <https://press.uchicago.edu/ucp/books/book/chicago/E/bo5961634.html>.
22. Wolfe KH, Li WH, Sharp PM. Rates of nucleotide substitution vary greatly among plant mitochondrial, chloroplast, and nuclear DNAs. *Proc Natl Acad Sci U S A*. 1987; 84(24): 9054-8. doi: <https://doi.org/10.1073/pnas.84.24.9054>.
23. Wang X, Cheng X, Zhou W, Zhang XX, Hu Y, Chen XY, Hu XS. Assessing the ecological and evolutionary processes underlying cytonuclear interactions. *Scientia Sinica Vitae*. 2019; 49(8): 951-64. doi: <https://doi.org/10.1360/SSV-2019-0049>.
24. Bremer B, Manen JF. Phylogeny and classification of the subfamily Rubioideae (Rubiaceae). *Pl Syst Evol*. 2000; 225(1): 43-72. doi: <https://doi.org/10.1007/BF00985458>.
25. Andreasen K, Bremer B. Combined phylogenetic analysis in the Rubiaceae-Ixoroideae: morphology, nuclear and chloroplast DNA data. *Am J Bot*. 2000; 87(11): 1731-48. doi: <https://doi.org/10.2307/2656750>.
26. Andersson L, Antonelli A. Phylogeny of the tribe Cinchoneae (Rubiaceae), its position in Cinchonoideae, and description of a new genus, *Ciliosemina*. *Taxon*. 2005; 54(1): 17-28. doi: <https://doi.org/10.2307/25065412>.